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## Synthesis, activity, metabolic stability, and pharmacokinetics of glucocorticoid receptor modulator—statin hybrids

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Abstract—The synthesis, activity, metabolic stability, and pharmacokinetics of steroidal and nonsteroidal glucocorticoid receptor modulator–statin hybrids is reported. Potent steroidal antagonist–statin hybrids like 22 (h-GR binding  $IC_{50} = 7 \text{ nM}$ ) and nonsteroidal modulator hybrids like 16 (h-GR binding  $IC_{50} = 2 \text{ nM}$ ) were discovered. Appending a 'statin'-like diol-acid group to the modulators dramatically improved metabolic stability (and in some cases hepatocyte activity), but did not impart hepatoselectivity. © 2004 Elsevier Ltd. All rights reserved.

Metformin has become a cornerstone of antidiabetic treatment both as a mono-therapy and in combination with other agents. Although its precise mechanism of action is still under investigation, its predominant activity is inhibition of hepatic glucose production (HGP). Its therapeutic and commercial success has stimulated research on multiple targets whose regulation may moderate HGP.<sup>2</sup> Antagonists of the glucocorticoid receptor (GR) have been explored as potential antidiabetic agents that inhibit HGP.3 Because of the possibility of undesired effects due to extrahepatic GR antagonist exposure, attempts have been made to target them to the liver.4 Steroidal GR antagonists, like mifepristone (RU-486) (1), are well known members of this class (Fig. 1). However, they are unselective and show significant progesterone receptor (PR) activity leading to their abortive properties. Recently, several classes of nonsteroidal GR ligands, including a series of GR selective modulators from our laboratory have been identified.5,6 Both the steroidal and nonsteroidal GR antagonists are typically widely distributed after oral

Figure 1. GR ligands and HMG CoA reductase inhibitors.

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dosing potentially leading to both hepatic and systemic effects. We have sought to alter the pharmacokinetics of

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GR antagonist mifepristone (1) and the GR selective nonsteroidal modulator 2 to increase their hepatic selectivity.

One example of liver selective small molecule therapeutics is the statin class of 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors. These compounds, which prevent coronary heart disease by lowering lowdensity lipoprotein cholesterol levels, show preferential distribution and action in the liver. However, some potentially beneficial extrahepatic effects have also been studied.<sup>8</sup> Lovastatin (3) and pravastatin (4) are two statins that have polar sidechains that structurally represent the class. Inactive hydroxylactones, like 3, upon ingestion and absorption, undergo lactone hydrolysis to the active diol-acids. Frequently the lipophilic lactones are able to passively cross membranes more efficiently than the polar diol-acids. Several diol-acids, like pravastatin (4), are passively and actively taken up by transporters (frequently the organic ion transporter) into hepatocytes. This increases their hepatoselectivity. Pravastatin (4) is orally absorbed without the necessity of administering the corresponding lactone.

We prepared hydroxylactone GR antagonist/modulator-hybrids 5 and diol-acid GR antagonists/modulatorhybrids 6 in order to liver target GR antagonists or modulators (Fig. 2). In the case of hybrids with the aniline bearing site of mifepristone (1), the GR binding activity was expected to be similar to 1 because the appended groups would project outside of the ligandbinding domain of the GR. 10 Reported structure activity studies also indicated large groups could be linked to this site without large reductions in potency. 11 The activity of the hybrids of the nonsteroidal GR modulator 2 was less predictable. The site of attachment and linker would need to be experimentally determined. If these hybrids target the GR modulators to the liver they could be optimized for dual lipid-lowering activity. Alternatively, HMG CoA activity could be potentially removed by altering the diol stereochemistry.

Two sites in the nonsteroidal modulators tolerated large polar substituents. At these positions, GR modulator-statin hybrids were prepared using an aryl ether linker. The core of one modulator was prepared from hydro-quinone 7 (Scheme 1). Allylation followed by a nucleophilic aromatic substitution reaction with 4-

**5** GR Modulator Lactone Hybrid **6** GR Modulator Diol-Acid Hybrid

Figure 2. GR ligand-statin hybrids.

Scheme 1. Reagents and conditions: (a) i. allyl iodide, K<sub>2</sub>CO<sub>3</sub>, acetone, 40 °C, 12 h, 41%, ii. 4-fluorobenzaldehyde, K<sub>2</sub>CO<sub>3</sub>, DMF, 100 °C, 12 h, 81%; (b) 2-methyl-3-nitroaniline, AcOH, DCE, rt, 4 h, Na(OAc)<sub>3</sub>BH, 12 h, 85%; (c) benzylbromide, *i*-Pr<sub>2</sub>NEt, DMF, 90 °C, 12 h, 94%; (d) Pd(PPh<sub>3</sub>)<sub>4</sub>, PhSiH<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, rt, 1 h, 95%.

fluorobenzaldehyde provided aldehyde **8**. Reductive amination with 2-methyl-3-nitroaniline followed by alkylation with benzyl bromide gave allyl ether **10**. Deprotection using tetrakis(triphenylphosphine)palladium[0] in the presence of phenylsilane yielded phenol **11**. 12

The key portion of the statins was appended from a known precursor via precedented alkylation chemistry (Scheme 2).<sup>13</sup> Phenol **11** and mesylate **12** reacted to give ether **13** in 62% yield. Conversion of nitroarene **13** into methylsulfonamide **14**, followed by deprotection, gave the diol-acid **16** in good yield. Lactone **17** was prepared from *t*-butyl ester **15** by treatment with trifluoroacetic acid. Steroidal GR antagonist–statin hybrids **21–24** were prepared from a known intermediate using a similar strategy.<sup>14</sup>

The antagonists and modulators were assayed in a h-GR binding assay, monitoring for the displacement of radiolabeled dexamethasone. Functional activity was assessed in a GR alkaline phosphatase reporter whole cell assay (GRAF). <sup>6a</sup> A second assay measuring the blockade of dexamethasone-induced activity of tyrosine amino transferase (TAT) in freshly isolated rat hepatocytes was also employed. Since rat hepatocytes have transporters (the organic ion transporter among others), it was hoped diol-acid hybrids would show improved activity in this assay. Weaker activity might also be observed in the GRAF assay if the statin substituents lowered cell permeability.

The nonsteroidal GR modulator **2** binds potently to h-GR ( $IC_{50} = 2.7 \text{ nM}$ ) (Table 1). It also has excellent selectivity over other nuclear hormone receptors (>100×), including human progesterone receptor (h-PR), mineralocorticoid (h-MR), androgen receptor (h-

**Scheme 2.** Reagents and conditions: (a)  $K_2CO_3$ , 18-crown-6, DMSO, 80 °C, 18 h, 62%; (b) Fe, NH<sub>4</sub>Cl, EtOH, H<sub>2</sub>O, 80 °C, 1 h; MsCl, pyr, rt, 1 h, 84%; (c) 3 N HCl, THF, EtOH, rt, 12 h, 97%; (d) NaOH, EtOH, rt, 2.5 h, 72%; (e) TFA, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C  $\rightarrow$  rt, 2 h, 82%.

Table 1. GR binding and functional assay results for nonsteroidal modulators 2 and 16-20

Compds	$R_1$	$\mathbf{R}_2$	R <sub>3</sub>	$R_4$	h-GR Binding IC <sub>50</sub> , nM <sup>a</sup>	GRAF IC <sub>50</sub> , nM <sup>a</sup>	Rat TAT IC <sub>50</sub> , μM <sup>a</sup>
2	Н	Н	Н	Н	2.7	240	>30
16	HO HO OH OH	Н	Н	Н	2.1	300	>30
17	HO Ost	Н	Н	Н	2.1	440	>30
18	Н	Н	F	F	21	385	>30
19	Н	HO OH OH	F	F	15	760	>30
20	Н	HO O'S	F	F	25	670	>30

<sup>&</sup>lt;sup>a</sup> Values are the geometric means of two experiments (na = not active, nd = not determined).

AR), and estrogen receptor (ER $_{\alpha}$  and  $_{\beta}$ ) (data not shown). However, sulfonamide 2 has modest functional

activity in the GRAF assay and is inactive in the rat hepatocyte TAT assay. para-Substituted hybrids

diol-acid **16** and lactone **17** have similar activity to the parent structure and are equally selective for h-GR. Diol-acid **16** shows little species selectivity (r-GR binding  $IC_{50} = 2.6 \,\text{nM}$ ). *meta*-Substituted hybrids of the h-GR selective difluoride **18** were also prepared. Although less potent, these hybrids (**19** and **20**) also maintained the activity of the parent **18**. Interestingly, the diol-acid **16** and lactone **17** show similar activity, as do the other pair **19** and **20**. Although potent binding statin hybrids were identified their hepatocyte functional activity did not improve.

Steroidal GR antagonists-statin hybrids were also evaluated (Table 2). The steroidal core is less selective due to its potent antagonism of h-PR. A shorter monohydroxyacid 21 showed diminished activity relative to mifepristone (1). However, the longer chain diolacid 22 and corresponding lactone 23 retain similar h-GR binding potency to the parent 1. Aniline hybrids 22 and 23 also have potent functional antagonist activity (GRAF  $IC_{50} = 18$  and 23 nM, respectively). Ether hybrid diol-acid 24 has potent binding activity but has significantly weaker GRAF activity, relative to mifepristone (1) and closely related aniline 21, indicating it is probably not cell permeable. Interestingly, all the compounds show good potency in the rat hepatocyte TAT assay. This suggests diol-acid 24 and the other steroid hybrids are actively transported into hepatocytes.

The metabolic stability of representative GR modulator and antagonist-statin hybrids were evaluated in rat and human microsomes and hepatocytes. For the lactone 17, the appearance of the corresponding diol-acid 16 was also monitored. Diol-acid 16 was metabolically stable when exposed to rat, or human, liver microsomes (incubation period = 60 min). The parent dibenzylaniline 2 is rapidly metabolized in microsomal incubations indicating the diol-acid group stabilizes the core structure. This is a desirable property for a liver targeting group. Lactone 17 under identical conditions converted to the stable diol-acid 16 during the incubation period. Diol-acid 16 underwent metabolism by both rat and human hepatocytes. In rat hepatocytes, metabolism occurred more extensively consuming all but 5%, whereas 60% of the parent remained at the end of the in human hepatocytes (6h studies). The identity of the metabolites was not determined but they are less polar than the diol-acid 16. This led us to speculate that they are dehydration products. During these experiments it was also noted that lactone 17 converts to acid 16 in buffer (half life not determined). This may also be the cause of their similar activity in the in vitro assays.

Pharmacokinetic studies at 5 mpk in Sprague–Dawley rats of open chain acid **16** showed the compound was cleared rapidly ( $Cl_p = 5.2 \text{ L/h kg}$ ), had a high volume of distribution ( $V_B = 6.2 \text{ L/kg}$ ), modest AUC (1.0 µg h/mL)

Table 2. GR binding and functional assay results for steroidal antagonists 1 and 21-25

Compds	R	h-GR binding IC <sub>50</sub> , nM <sup>a</sup>	h-PR Binding IC <sub>50</sub> , nM	GRAF IC <sub>50</sub> , nM <sup>a</sup>	Rat TAT IC <sub>50</sub> , μM <sup>a</sup>
1	NMe <sub>2</sub>	1.1	2.9	4.8	0.27
21	HO OH	37	200	76	0.22
22	HO OH OH	6.8	32	18	0.13
23	HO Note	2.3	3.8	23	0.14
24	HO OH OH	9.2	47	2900	0.56

<sup>&</sup>lt;sup>a</sup> Values are the geometric means of two experiments (na = not active, nd = not determined).

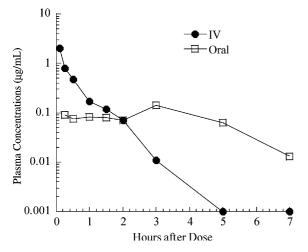


Figure 3. Rat pharmacokinetics of GR modulator-statin hybrid 16.

and a short half life  $(t_{1/2} = 48 \,\mathrm{min})$  after iv dosing. Bioavailability was moderate (F = 54%) (Fig. 3). Although the high clearance may be consistent with liver uptake, liver levels at  $t = 7 \,\mathrm{h}$  were low  $(0.024 \,\mu\mathrm{g/g})$  in the iv animals and the orally treated animals were not appreciably higher  $(0.1 \,\mu\mathrm{g/mg})$ . Diol-acid 19 gave a similar profile. Oral administration of the corresponding lactone 20 provided nearly identical blood and liver levels of diol-acid 19 when compared with oral dosing of 19 itself. The data is consistent with lactone hydrolysis prior to absorption. No lactone 20 was detected in either plasma or liver.

Steroidal hybrid **24** had a similar profile to the nonsteroid **16**. Steroidal diol-acid **24** was stable in rat and human liver microsomes (>90% remaining after 60 min). Good stability was also observed in rat (75% remaining) and human (>95% remaining) hepatocytes after a 6 h incubation. As with the nonsteroidal diol-acids, this profile represents a substantial improvement over the rapid metabolism of mifepristone (1). Oral/iv pharmacokinetic studies at 5 mg/kg in Sprague–Dawley rats indicated steroidal antagonist **24** had a high iv clearance consistent with liver uptake. However, low bioavailability (<5%) and liver levels (<5  $\mu$ g/g) after iv or oral dosing were observed.

Nonsteroidal GR selective modulator-statin hybrids and steroidal GR antagonist-statin hybrids were prepared that have similar nuclear hormone activity as their potent parent compounds. The dihydroxyhexanoic acid side chains improved the metabolic stability of both the steroidal antagonist and nonsteroidal modulator. It also improved the cellular activity of the steroid hybrids in hepatocytes. These are valuable properties for a polar low molecular weight group to confer. However, neither class of hybrids achieved the necessary liver levels (or liver selectivity) to provide sufficient GR blockade to lower blood glucose levels. The successful realization of liver targeting for these series with other structural motifs will be reported in future publications. <sup>11</sup>

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## References and notes

- 1. Cusi, K.; DeFronzo, R. A. *Diabetes Rev.* **1998**, *6*, 89–131.
- (a) Kurukulasuriya, R.; Link, J. T.; Madar, D. J.; Pei, Z.; Rohde, J. J.; Richards, S. J.; Souers, A. J.; Szczepankiewica, B. G. Curr. Med. Chem. 2003, 10, 99–122; (b) Kurukulasuriya, R.; Link, J. T.; Madar, D. J.; Pei, Z.; Richards, S. J.; Rohde, J. J.; Souers, A. J.; Szczepankiewica, B. G. Curr. Med. Chem. 2003, 10, 123–154.
- Friedman, J. E.; Sun, Y.; Ishizuka, T.; Farrell, C. J.; McCormack, S. E.; Herron, L. M.; Hakimi, P.; Lechner, P.; Yun, J. S. *J. Biol. Chem.* 1997, 272, 31475–31481.
- (a) Apelqvist, T.; Wu, J.; Koehler, K. F. WO00058337, 2000; (b) Sorensen, B.; Link, J. T.; von Geldern, T.; Emery, M.; Wang, J.; Hickman, B.; Grynfarb, M.; Goos-Nilsson, A.; Carroll, S. *Bioorg. Med. Chem. Lett.* 2003, 13, 2307–2310.
- (a) Link, J. T.; Sorensen, B. K.; Patel, J. R.; Arendsen, D. L.; Li, G. WO 064550, 2002; (b) Link, J. T.; Sorensen, B. K.; Patel, J.; Emery, M.; Grynfarb, M.; Goos-Nilsson, A. Bioorg. Med. Chem. Lett. 2004, 14, 2209–2212; (c) Link, J. T.; Sorensen, B. K.; Patel, J.; Arendsen, D.; Li, G.; Swanson, S.; Nguyen, B.; Emery, M.; Grynfarb, M.; Goos-Nilsson, A. Bioorg. Med. Chem. Lett. 2004, 14, in press. doi:10.1016/j.bmcl.2004.06.023.
- (a) Apelqvist, T. WO 63976, 1999; (b) Kym, P.; Lane, B.; Pratt, J.; von Geldern, T.; Winn, M.; Brenneman, J.; Patel, J.; Arendsen, D.; Akritopoulou-Zanze, I. U.S. Patent 6,329,534 B1, 2001; (c) Miner, J. N.; Tyree, C.; Hu, J.; Berger, E.; Marschke, K.; Nakane, M.; Coghlan, M. J.; Clemm, D.; Lane, B.; Rosen, J. Mol. Endocrinol. 2002, 17, 117–127; (d) Morgan, B. P.; Swick, A. G.; Hargrove, D. M.; LaFlamme, J. A.; Moynihan, M. S.; Carroll, R. S.; Martin, K. A.; Lee, E.; Decosta, D.; Bordner, J. J. Med. Chem. 2002, 45, 2417–2424; (e) Grese, T. A.; Jadhav, P. K.; Neel, D. A.; Steinberg, M. I.; Lander, P. A. WO 078394, 2003; (f) Amjad, A.; Balkovec, J. M.; Graham, D. W.; Thompson, C. F.; Nazia, Q. WO 086294, 2003.
- 7. (a) Hamelin, B. A.; Turgeon, J. Trends Pharm. Sci. 1998, 19, 26–37; (b) Corsini, A.; Bellosta, S.; Baetta, R.; Fumagalli, R.; Paoletti, R.; Bernini, F. Pharm. Ther. 1999, 84, 413–428; (c) Hatanaka, T. Clin. Pharmacokinet. 2000, 39, 397–412; (d) McTaggart, F.; Buckett, L.; Davidson, R.; Holdgate, G.; McCormick, A.; Schneck, D.; Smith, G.; Warwick, M. Am. J. Cardiol. 2001, 87, 28D–32B; (e) Nezasa, K.; Higaki, K.; Matsumura, T.; Inazawa, K.; Hasegawa, H.; Nakano, M.; Koike, M. Drug Metab. Dispos. 2002, 30, 1158–1163.
- 8. Mundy, G.; Garrett, R.; Harris, S.; Chan, J.; Chen, D.; Rossini, G.; Boyce, B.; Zhao, M.; Gutierrez, G. *Science* **1999**, *286*, 1946–1949.
- Nakai, D.; Nakagomi, R.; Furuta, Y.; Tokui, T.; Abe, T.; Ikeda, T.; Nishimura, K. J. Pharm. Exp. Ther. 2001, 297, 861–897.
- 10. Kauppi, B.; Jakob, C.; Färnegårdh, M.; Yang, J.; Ahola, H.; Alarcon, M.; Calles, K.; Engström, O.; Harlan, J.;

- Muchmore, S.; Ramqvist, A.-K.; Thorell, S.; Öhman, L.; Greer, J.; Gustaffson, J.-A.; Carlstedt-Duke, J.; Carlquist, M. *J. Biol. Chem.* **2003**, *278*, 22748–22754.
- von Geldern, T.; Tu, N.; Kym, P. R.; Link, J. T.; Jae, H.; Lai, C.; Apelqvist, T.; Rhonnstadt, P.; Hagberg, L.; Koehler, K.; Grynfarb, M.; Goos-Nilsson, A.; Sandberg, J.; Österlund, M.; Wang, J.; Fung, S.; Wilcox, D.; Nguyen, P.; Jakob, C.; Hutchins, C.; Färnegårdh, M.; Kauppi, B.; Öhman, L.; Jacobsen, P. B. J. Med. Chem., in press.
- 12. Dessolin, M.; Guillerez, M.-G.; Thieriet, N.; Guibé, F.; Loffet, A. *Tetrahedron Lett.* **1995**, *36*, 5741–5744.
- Jendralla, H.; Granzer, E.; Kerekjarto, B. V.; Krause, R.; Schacht, U.; Baader, E.; Bartmann, W.; Beck, G.; Bergmann, A.; Kesseler, K.; Wess, G.; Chen, L.-J.; Granata, S.; Herchen, J.; Kleine, H.; Schüssler, H.; Wagner, K. J. Med. Chem. 1991, 34, 2962–2983.
- (a) Bélanger, A.; Philibert, D.; Teutsch, G. Steroids 1981,
   37, 361–382; (b) Acosta, K.; Cessac, J. W.; Rao, N.; Kim,
   H. K. J. Chem. Soc., Chem. Commun. 1994, 1985–1986.